Solid-Phase Organometallic Synthesis

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Abstract: A solid-phase synthesis approach for a class of molybdenum carbonyl complexes has been developed. The system can be used to perform metalcomplexation, ligand substitution reactions and oxidative eliminations on the solid phase and to cleave the final complexes under mild and selective conditions. Comparison is made to corresponding soluble complexes and liquid-phase reactions.

Introduction

Solid-phase organic synthesis (SPOS, Scheme 1) has become an increasingly useful method in synthetic organic and biological (especially pharmaceutical) chemistry. Generally, the compounds synthesised on the solid phase are *organic*



Scheme 1. Principle of solid-phase synthesis.

molecules, while "*organometallic* compounds are usually generated on insoluble supports as synthetic intermediates only, and not as target molecules".^[1] A few exceptions are the solid-phase syntheses of metal complex functionalised oligo-nucleotides and oligodesoxynucleotides using standard DNA/RNA syntheses,^[2] metal complex functionalised peptides using standard peptide chemistry,^[3] and an attempt to prepare a 2,2'-bipyridine – dichloroplatinum derivative.^[4]

Metal complexes have found applications as linkers^[5] and as cleavage reagents^[1, 6] in solid-phase chemistry, but the most important and most frequent use of metal complexes in this

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context is catalysis. In this case the metal complex remains bound to the solid support.

Coordination and organometallic chemistry on the solid phase has not yet been extensively explored, usually only the catalyst performance itself has been studied. Elementary reaction steps such as ligand substitution have not yet been investigated in detail under solid-phase reaction conditions despite the fact that in some cases catalytic results, activity and (enantio-) selectivity, are strikingly different when compared with the homogeneous case.^[7]

In solid-phase chemistry the support can render increased stability to reactive intermediates by blocking bimolecular reaction steps^[8] and can even change reaction pathways from ionic to radical mechanisms.^[9] With this background in mind it appears useful and necessary to investigate elementary *inorganic* reaction steps performed on the insoluble support and to compare these results with conventional liquid-phase inorganic chemistry.

For this purpose a system has been designed, which makes it possible to perform standard reactions of coordination and organometallic chemistry under solid-phase reaction conditions and—for ease of reaction control and characterisation to cleave the metal complexes from the solid phase.

The target molecules investigated herein are molybdenum carbonyl complexes, which have several important properties: 1) stability, 2) rich coordination chemistry, 3) CO ligands that serve as a sensitive analytical probe (IR), and 4) an NMR-active metal nucleus. In addition to these intrinsic properties molybdenum carbonyl complexes with nitrogen donor ligands have found applications as catalysts in allylic substitution reactions,^[10] as cocatalysts for olefin metathesis,^[7c] and they have proven useful for luminescence and electron transfer studies.^[11]

Results and Discussion

Solution chemistry, ligand synthesis: The ligands employed are the bidentate nitrogen base **1a** with a phenolic hydroxy

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group for attachment on the solid support and the trimethylsilyl (TMS)-protected analogue **2a** as a model for the immobilised compounds (Scheme 2).



Scheme 2. Syntheses of ligands 1a and 2a.

Schiff base condensation of p-aminophenol and pyridine-2carbaldehyde gives the chelating ligand 1a and silylation of 1awith trimethylsilyl chloride and triethylamine as the base resulted in the model analogue 2a (Scheme 2). Both compounds are fully characterised (see Experimental Section).

Solution chemistry, model complexes: Common synthetic approaches to complexes of the type $(N N')Mo(CO)_3L$ (N N' = symmetrical or asymmetrical bidentate nitrogen donor ligand, L = monodentate ligand) are depicted in Scheme 3.^[12] All of them use high temperature or give only the tetracarbonyl compounds (L=CO), which then have to be converted into the desired tricarbonyl compounds—again using high temperature and rather harsh oxidative reaction conditions.

For a successful use in solid-phase chemistry with polystyrene resins, such harsh conditions have to be avoided as the molybdenum precursors can react with the aromatic residues of the support. Therefore a selective high-yield route using mild reaction conditions was elaborated with $[(CH_3CN)_3Mo (CO)_3]$ as the "Mo(CO)₃" source (Scheme 3, lower right).



Scheme 3. Synthetic approaches to complexes of the type [(N^N')-Mo(CO)_3L].

Reaction of **1a/2a** with tris(acetonitrile)molybdenumtricarbonyl in acetonitrile at room temperature rapidly and cleanly gives the intensely blue-coloured complexes **1b/2b** in good to excellent yields (Scheme 4).

When the same reactions are performed in CH_2Cl_2 the tetracarbonyl complexes 1c/2c are formed as shown by IR, UV/Vis/NIR, and NMR spectroscopy (Scheme 5). The latter reaction is always observed when dissolving complexes 1b/2b in CH_2Cl_2 . CO scrambling promoted by dichloromethane has been observed previously.^[12a]

UV/Vis/NIR spectroscopy indicates the stoichiometric formation of **1c/2c** (final absorbance at 539 nm) plus an unidentified side product without carbonyl groups (as shown by IR spectroscopy; isosbestic point at 1821 cm⁻¹; see Supporting Information). The absence of well-defined isosbestic points (Figure 1, top) additionally indicates a complex reaction. From the absorbance changes at 500 and 539 nm a pseudo-first-order rate constant for the formation of **2c** can be obtained $k = 0.9 \times 10^{-4} \text{ s}^{-1}$ (500 nm; R² = 0.9993) and $k = 1.1 \times 10^{-4} \text{ s}^{-1}$ (539 nm; R² = 0.9978) in good agreement (Figure 1, bottom). For complex **1b** this reaction proceeds even faster, precluding a reliable rate constant determination but giving the same results with respect to the stoichiometry of the reaction.





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Scheme 5. Reaction of 1b/2b to 1c/2c in CH₂Cl₂.



Figure 1. UV/Vis/NIR spectra of complex 2b in CH₂Cl₂ (top) and the course for the formation of 2c as a function of time (bottom).

Table 1. ¹H NMR data for complexes 1b-1f and 2b-2e in CD_2Cl_2 .^[a]

The substitutional lability of the acetonitrile ligand in 1b and 2b has been utilised for ligand-exchange reactions with carbon monoxide, *tert*-butyl isonitrile, and triphenylphosphane giving the corresponding complexes 1c-e and 2c-e, respectively (Scheme 4). Reaction of 1c with phenyltin trichloride as an example of an oxidative elimination gives the seven-coordinate molybdenum complex 1f, while starting from 2c does not produce the corresponding complex 2f but rather 1f, as the trimethylsilyl group is cleaved under these conditions (Scheme 6). All complexes have been thoroughly characterised by spectroscopic and analytical methods (Tables 1-3).

For **2e** the ¹³C resonance signals of the CO groups are split into doublets by coupling to the ³¹P nucleus. Coupling of the phosphorus atom is also observed to H⁷ (${}^{4}J_{\rm PH}$ =2.6 Hz; confirmed by a ³¹P-¹H COSY experiment) and to C⁷ (${}^{3}J_{\rm PC}$ = 3.0 Hz) of the Schiff base ligand. Complexes **1e**/**2e** each show



Scheme 6. Synthesis of 1 f.

a singlet in the ³¹P NMR spectrum, confirming the presence of only one isomer.

The ⁹⁵Mo NMR signals (Table 2) appear in the expected region.^[12f, 13] The different ligands L induce the expected high field shift of the ⁹⁵Mo resonances as well as of the ¹³C

	$1b^{[b]}$	1c	1 d	1e	1 f ^[c]	$2b^{[b]}$	2 c	2 d	2 e
H^2/H^6	7.56	7.55	7.57	7.37	7.41	7.59	7.54	7.58	7.39
	(d, 8.5 Hz)	(d, 8.7 Hz)	(d, 8.2 Hz)	(d, 8.2 Hz)	(d, 8.4 Hz)	(d, 8.8 Hz)	(d, 8.1 Hz)	(d, 8.5 Hz)	(d, 8.4 Hz)
H^{3}/H^{5}	6.91	7.00	6.99	6.92	6.87	6.97	7.01	6.98	6.94
	(d, 8.5 Hz)	(d, 8.7 Hz)	(d, 8.2 Hz)	(d, 8.2 Hz)	(d, 8.4 Hz)	(d, 8.8 Hz)	(d, 8.1 Hz)	(d, 8.5 Hz)	(d, 8.4 Hz)
H ^{7[e]}	8.65 (s)	8.57 (s)	8.56 (s)	8.18 (bs)	8.64 (s)	8.68 (s)	8.58 (s)	8.57 (s)	8.18 (d, ^[d])
$H^{9[e]}$	7.9-8.1	7.92	7.89	7.44	8.09	7.94	7.9-8.0	7.9-8.0	7.45
	(m)	(d, 7.4 Hz)	(d, 8.0 Hz)	(d, 8.0 Hz)	(d, 7.2 Hz)	(d, 7.2 Hz)	(m)	(m)	(d, 8.0 Hz)
H^{10}		8.02 (pt)	7.91 (pt)	7.68 (pt)	8.21 (pt)	8.03 (pt)			7.68 (pt)
H^{11}	7.50 (pt)	7.56 (pt)	7.42 (pt)	7.01 (pt)	7.78 (pt)	7.50 (pt)	7.52 (pt)	7.42 (pt)	7.03 (pt)
H^{12}	9.03	9.21	9.23	8.76	9.10	9.03	9.20	9.23	8.77
	(d, 5.0 Hz)	(d, 4.8 Hz)	(d, 4.6 Hz)	(d, 4.4 Hz)	(d, 4.8 Hz)	(d, 5.2 Hz)	(d, 3.8 Hz)	(d, 4.8 Hz)	(d, 4.8 Hz)
OH/OTMS	7.1 (bs)	n. o.	n. o.	n. o.	8.87 (s)	0.31 (s)	0.38 (s)	0.38 (s)	0.42 (s)
ligand L	1.92	_	1.33	7.19-7.23	7.38	1.93	-	1.33	7.1-7.3
-	(CH ₃ , s)		(CH ₃ , s, 9H)	(m, 15H)	$(C^{o} + C^{p}, m, 3H)$ 8.00 $(C^{m}, m, 2H)$	(CH ₃ , s)		(CH ₃ , s, 9H)	(m, 15H)

[a] pt = pseudo-triplet; bs = broad singlet; n. o. = not observed. [b] In CD₃CN. [c] In $[D_8]$ THF. [d] ${}^{4}J_{PH} = 2.6$ Hz. [e] The cisoid conformation of the Schiff base ligand in the complexes is evidenced in solution by the observation of an NOE between H⁷ and H⁹ which is absent in the spectra of the free ligands.

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10002. C, $100, 1, and 50 10000 complexes 10, 11, and 20-20 in CD2Cl2.$	Table 2.	¹³ C, ⁹⁵ M	o, ³¹ P, a	and ¹¹⁹ Sn	NMR	data for	complexes	1b, 1f	f, and 2b -	-2e in	CD ₂ Cl	[a]
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	1 b ^[b]	1 f ^[c]	2 b ^[b]	2b ^[d]	2 c	2 d	2e
C1	145.5	149.8	146.8	145.9	146.4	146.9	146.6
C^{2}/C^{6}	124.7	124.4	124.6	123.6	123.8	123.8	124.4
C^{3}/C^{5}	116.4	116.7	121.0	120.4	120.8	120.5	120.5
C^4	155.8	154.1	155.8	155.1	154.8	155.0	154.3
C^7	162.6	166.6	163.2	160.6	160.9	159.2	158.6 (d, ${}^{3}J_{PC} = 3.0 \text{ Hz}$)
C ⁸	158.3	157.9	156.6	156.2	156.6	156.2	156.3
C ⁹	129.2	130.4	129.3	128.1	128.6	128.1	128.0
C^{10}	138.7	141.7	138.7	137.3	137.4	136.4	135.3
C ¹¹	127.6	130.0	127.8	126.6	126.8	125.9	125.7
C ¹²	152.7	154.3	152.8	152.3	153.4	152.5	152.3
OTMS	-	-	0.13	0.11	0.28	0.29	0.36
C ¹³ /C ¹⁵	228.9/	224.9 ^[e]	228.8/	228.8/	222.2/	227.2/	227.8 (d, ${}^{2}J_{PC} = 10.0 \text{ Hz})/$
	231.1	227.7 ^[e]	231.1	230.7	224.8	230.3	230.7 (d, ${}^{2}J_{\rm PC} = 8.5$ Hz)
C^{14}	219.0	215.1 ^[e]	218.8	217.0	203.6	210.4	213.5 (d, ${}^{2}J_{\rm PC} = 35.0 \rm Hz$)
ligand L		143.8 (C^{i})		1.95 (CH ₃)		$30.9 (CH_3)$	134.1 (C^{i} , d, ${}^{1}J_{PC} = 28.0 \text{ Hz}$)
		129.2 ($C^{\circ}, {}^{2}J_{\text{Sn}C} = 87 \text{ Hz}^{[f]}$)		116.9 (C_q)		56.5 (C_q)	133.4 (C° , d, ${}^{2}J_{P,C} = 12.5$ Hz)
		136.2 ($C^{\rm m}$, ${}^{3}J_{\rm sn C} = 61$ Hz ^[f])					128.4 ($C^{\rm m}$, d, ${}^{3}J_{\rm P,C} = 8.5$ Hz)
		$(C^{\rm p}, {}^{4}J_{\rm c_{rC}} = 18 \text{ Hz}^{[f]})$					129.3 (<i>C</i> ^p , s)
⁹⁵ Mo δ (Δv_{10} / Hz)	- 969 (50)	n. o. ^[g]	- 953 (65)	- 933 (80)	- 1160 (80)	- 1095 (130)	-1043 (80) ^[h]
$^{31}P \delta (\Delta v_{1/2}/Hz)$	_	_	_	_	_	_	39.4 (4)
¹¹⁹ Sn δ ($\Delta \nu_{1/2}$ / Hz)	_	30 (265)	_	-	_	_	_

[a] Assignments are based on the intensity, chemical shifts, and DEPT and 2D NMR experiments. [b] In CD₃CN. [c] In [D₈]THF. [d] Due to the long acquisition time the ¹³C and ⁹⁵Mo NMR spectra in CD₂Cl₂ additionally show signals of the tetracarbonyl complex **2c** but these are easily distinguished from those of **2b**. [e] At 193 K; not observed at 303 K. [f] ¹¹⁹Sn satellites. [g] n. o. = not observed. [h] ¹J_{Mo.P} = 138 Hz.

Table 3. IR, UV/Vis/NIR and CV data ($E_{1/2}$ vs. SCE) for complexes 1b-1f and 2b-1e in CH₂Cl₂.

	$ ilde{ u}_{ m CO} [m cm^{-1}] \ (m CH_2Cl_2/CsI)$	$ ilde{ u}_{ m CN}$ (CH ₂ Cl ₂ /CsI)	$ ilde{ u}_{ m OH}$ (CH ₂ Cl ₂ /CsI)	$egin{aligned} &\lambda_{\max} \ [nm] \ &(arepsilon_{\max} \ [M^{-1} ext{cm}^{-1}]) \end{aligned}$	$E_{_{1/2}} [\mathrm{mV}] \ (\Delta E_{_{1/2}} [\mathrm{mV}])$
1b	1909 (vs), 1835 (sh), 1794 (vs) ^[a] /	-	_	589 (4400) ^[a]	+185
	1898 (vs), 1833 (sh), 1782 (vs)		3230 (br)	[624 in CH ₂ Cl ₂]	(110, qrev.)
1c	2016 (m), 1910 (vs), 1889 (sh), 1839 (s)/	-	-	537 (4700)	+650
	2018 (m), 1898 (vs), 1876 (vs), 1835 (vs), 1814 (vs)		3550 (m)		(100, qrev.)
1 d	1917 (vs), 1833 (s), 1803 (s)/	2138 (s)/	-	618 (6740)	+250
	1935 (vs), 1919 (vs), 1867 (vs), 1763 (vs) ^[b]	2122 (s)	3434 (s)		(85, qrev.)
1e	1919 (vs), 1827 (s), 1800 (s)/			615 (6145)	+365
	1911 (vs), 1823 (vs), 1765 (vs)		3448 (m)		(120, qrev.)
1 f	2010 (vs), 1930 (m), 1907 (s)/	-	_ ` `	474 (2515)	- 850 mV
	2009 (vs), 1926 (m), 1905 (vs)		3444 (m)		(irrev.; +40 mV)
2b	1910 (vs), 1835 (w), 1794 (s) ^[a] /	-	-	592 (4340) ^[a]	+215
	1894 (vs), 1774 (vs)			[628 in CH ₂ Cl ₂]	(90, qrev.)
2c	2016 (m), 1910 (vs), 1889 (sh), 1839 (s)/	-	-	539 (5820)	+660
	2020 (m), 1911 (vs), 1866 (vs), 1830 (s), 1804 (vs)				(115, qrev.)
2 d	1917 (vs), 1834 (s), 1804 (s)/	2127 (s)/	-	619 (6890)	+270
	1912 (vs), 1830 (s), 1802 (s)	2129 (s)			(115, grev.)
2e	1918 (vs), 1827 (s), 1800 (s)	-	_	616 (6160)	+ 355
	1911 (vs), 1826 (s), 1795 (s)			. ,	(100, qrev.)

[a] In CH₃CN. [b] Splitting of CO absorptions in the solid state probably due to hydrogen bonding.

resonance signals of the carbonyl group *trans* to L according to their π -acceptor strength CO > CN*t*Bu > PPh₃ > CH₃CN.^[14] The correlation of ⁹⁵Mo and ¹³C (*trans* to L) chemical shifts is acceptably linear (R²=0.9584; see Supporting Information), while no linear correlation is observed between the ⁹⁵Mo shift and other ¹³CO resonance signals. Neither does there exist a linear correlation between δ (⁹⁵Mo) and the lowest energy absorption λ_{max} (Table 3), suggesting that the paramagnetic shielding term σ_{para} is not only influenced by the average electronic excitation energy ΔE but also by nephelauxetic contributions.^[12e, 15] This is not unexpected as very different ligands concerning electronic and steric properties are considered.^[16] The linewidth of the ⁹⁵Mo signal allows the detection of the scalar coupling to ³¹P in complex **2e**, giving a doublet with ¹J_{Mo,P} = 138 Hz similar to that of comparable complexes.^[12f, 13]

The seven-coordinate complex $1 f^{[17]}$ is fluxional at room temperature preventing the observation of ¹³C NMR signals

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of the carbonyl groups at 300 K, while at 193 K three resonance signals are observed. The resonance signal of the tin atom in **1 f** can be seen in the ¹¹⁹Sn NMR spectrum at δ = 30 both at 300 and 193 K. The coordination shift amounts to 334 ppm relative to phenyltin trichloride (δ = -304). No signal from the molybdenum nucleus could be observed in the ⁹⁵Mo NMR spectrum of **1 f** at 300 K or at 193 K.

The IR spectra in solution (Table 3) are consistent with a disturbed local C_{3v} symmetry of the *fac* complexes 1b,1d-f, **2b**, 2d-e (splitting of the "E" band) and with disturbed local C_{2v} symmetry of the *cis*-tetracarbonyl complexes 1c and 2c. For 1f only one set of absorption bands is observed indicting that only one isomer is formed in solution.^[17]

The electronic spectra of 1b-1e show metal-to-ligand charge transfer (MLCT) absorption bands in the 590–630 nm range (Table 3). According to the literature^[12] these bands can be assigned to the (ligand- π^*) \leftarrow (metal-d) transition. Substitution of CO by L shifts this band to lower energy, as the metal d-orbitals are more destabilised by L than by CO. For **1 f** the MLCT band^[17] is found at 474 nm.

The same argument holds for the interpretation of the cyclic voltammograms: the destabilisation of the metal d orbital is reflected in lower oxidation potentials for complexes with L \pm CO (Table 3). The seven-coordinate Mo^{II} complex **1f** is irreversibly reduced at -850 mV (vs. SCE) with a new oxidation wave appearing in the reverse scan at +40 mV (vs. SCE).

Reactivity studies in solution show that ligand exchange reactions proceed in a facile way (Scheme 7). The order of increasing lability $CNtBu < CO < PPh_3 < CH_3CN$ has been established by IR and NMR spectroscopy.



Scheme 7. Ligand-exchange reactions.

Solid-phase chemistry, resin functionalisation: Before starting to study organometallic reactions on the solid phase it is necessary to introduce a linker between the resin and the metal complex (see Scheme 1). This linker has to fulfil several requirements: 1) stability during all reactions performed on the solid phase and 2) mild cleavage conditions with reagents compatible with the final metal complex. One possible candidate is a linker based on a silyl ether^[18, 19] that is stable under basic and acidic reaction conditions but can be cleaved with fluoride ions, which are expected to be unreactive towards most metal complexes.

The polystyrene resin was functionalised with a silyl chloride group (Scheme 8) according to a literature procedure.^[18] Loading with the phenolic ligand **1a** is accomplished



Scheme 8. Synthesis of resin 3a.

under basic conditions (Scheme 8). The yellow ligand modified resin **3a** was studied by gel-phase ¹³C NMR, IR, diffuse reflectance spectroscopy, and mass spectrometry to characterise the attached ligand and by differential scanning calorimetry, thermogravimetry, and studies of the swelling properties to characterise the bulk material. Quantitative loading with ligand **1a** was determined by elemental analysis (nitrogen content; 0.56 mmol **1a** per gram polymer) and metal ion uptake (CoCl₂ in THF; measured spectrophotometrically at 590 and 683 nm; 0.52 mmol g⁻¹). The (within error) identical results obtained by both methods indicate that all ligand sites within the polymer are accessible for metal complexation (in THF). The loading of about 0.5 mmol g⁻¹ corresponds to a statistical substitution of every tenth styrene residue with the ligand **1a**.

The swelling properties of polystyrene resins have been shown to be crucial for rapid and complete transformations. The most suitable solvents (Table 4) for resin **3a** were CH₂Cl₂, THF, and toluene (2.5 mLg⁻¹) followed by *N*,*N*-dimethylacetamide (DMA) (2.2 mLg⁻¹), ethyl acetate, and CH₃CN (2.0 mLg⁻¹). In contrast to polystyrene (with 2% divinylbenzene (DVB)) the swelling of **3a** is much poorer, probably because of additional cross-linking during the reaction with Cl₂Si(*i*Pr)₂ (Scheme 8). It has been proposed that swelling values greater than 4 mLg⁻¹ indicate good and values of 2.0– 4.0 mLg⁻¹ indicate only moderate swelling properties.^[20]

Although swelling of **3a** is rather poor, a gelphase ¹³C NMR spectrum could be obtained in $[D_8]$ THF, which only shows signals of the linker group and the attached ligand (Table 4) as these residues are the most mobile groups.

The covalent attachment of the linker and the ligand is also confirmed by EI mass spectrometry^[18] as characteristic signals of styrene fragments containing the linker and the ligand are observed, for example $[C_8H_7Si(iPr)_2OL]^+$ at m/z 415 (see Experimental Section).

The IR spectrum of 3a (as CsI disk) shows signals in addition to the signals of the polystyrene backbone due to the

Color		3a vellow	3b	3c	3d	3e	3f
		yenow	01dc			blue – green	
DRS-UV (PTFE)	$\lambda_{\rm max}$ [nm]	396	407,775	404,604	404,681	412,681	449, 533 (sh)
IR (CsI)	CH _{imine}	2726 (w)	2726 (w)	2727 (w)	2728 (w)	2722 (w)	2728 (w)
$\tilde{\nu}$	C=N _{imine}	1627 (m)	1627 (m)	1627 (m)	1627 (m)	n. o.	1637 (m)
$[cm^{-1}]$	pyridine	1584 (m)	1584 (m)	1584 (m)	1586 (m)	1586 (sh)	1584 (sh)
	CO	-		2013 (s)		[a]	
		-	1911 (vs)	1909 (vs)	1921 (vs)	1911 (vs)	2008 (s)
		-	1850 (s)	1896 (sh)	1848 (s)	1847 (s)	1930 (s)
		-	1811 (s)	1851 (s)	1821 (s)	1813 (s)	1912 (s)
	CNtBu	-	-	-	2112 (m)	-	-
¹³ C NMR	Si-CHCH ₃	18	[d]	18	18	18	16
δ ([D ₈]THF)	C^2 , C^3 , C^5 , C^6 , C^{11}	121-125		120-125	120-126	121-123	br
	C^7	158		155	159	n. o.	n. o.
	C^9	129		129	129	129	130
	C^{10}	136		137	138	138	138
	C^{12}	151		149	n. o.	n. o.	n. o.
	ligand	_		-	31 (CH ₃)	126, 129, 134	126, 130, 138
$^{31}P \delta (\Delta v_{1/2})$		_	-	-	-	40 (1000)	_
¹¹⁹ Sn δ ($\Delta v_{1/2}$)		_	-	_	-	-	50 (9140) ^[e]
DSC	$T [^{\circ}C]^{[b]}$		382 (sh)	385 (sh)	381 (sh)	381 (sh)	
	$[kJg^{-1}]$	434(-0.48)	421(-0.23)	421(-0.33)	422(-0.35)	421(-0.33)	425(-0.34)
TG	$T [^{\circ}C]^{[c]}$		117 (6)	111 (3)	104 (4)	119 (3)	113 (6)
	(weight		323 (25)	331 (25)	343 (32)	339 (27)	344 (40)
	loss/%)	358 (59)	390 (13)	399 (15)	401 (12)	402 (18)	394 (7)
	,		448 (19)	464 (19)	486 (15)	480 (15)	
		565 (33)	502 (30)	546 (31)	531 (30)	582 (30)	512 (40)
		8% residue	7;% residue	7% residue	7% residue	7% residue	7% residue
swelling	drv	1.6	1.6	1.6	1.5	1.5	1.5
$[mLg^{-1}]$	THF	2.5	2.3	2.2	2.3	2.2	2.2
	toluene	2.5	2.2	2.2	2.2	2.1	2.0

[a] In some preparations a band at 2013 cm^{-1} is observed indicating partial formation of the tetracarbonyl complex **3c**. [b] Peak maximum. [c] Inflection point. [d] Side reaction in THF prevents acquisition of NMR spectra. [e] At 203 K.

attached ligand at 2726 cm^{-1} (CH_{imine}), 1627 cm^{-1} (C=N_{imine}) and 1584 cm^{-1} (pyridine vibration). In the diffuse reflectance spectrum (measured in polytetrafluoroethylene, PTFE) a band around 400 nm is observed which is responsible for the yellow colour of the resin (Table 4, Figure 2).

The TG measurement of **3a** shows a two-step weight loss at 358 °C (59%) and 565 °C (33%); the remaining material (8%) being SiO₂.^[18] Differential scanning calorimetry (DSC) of **3a** shows an endothermic process at 434 °C (-0.48 kJ g^{-1}). Both measurements indicate that attachment of the linker and the ligand to the polystyrene has an influence on the thermal properties (depolymerisation) of the bulk polymer.^[18]

Solid-phase chemistry, resin complexation: Although CH_2Cl_2 was found to be a "good solvent" for swelling resin **3a**, performing the complexation of the polymer in this solvent leads to formation of the immobilised tetracarbonyl complex **3c** instead of the desired tricarbonyl complex **3b** (Scheme 4). This behaviour is analogous to the solution reaction (Scheme 5). The use of pure acetonitrile—a rather poor solvent for resin **3a**—leads to formation of the tricarbonyl complex **3b** but only with a low yield as shown by IR spectroscopy. This can be explained by the assumption that not all ligand sites are available when the resin is swollen in acetonitrile. Therefore a 1:1 mixture of toluene/acetonitrile was used (swelling 2.3 mLg⁻¹) for preparing the tricarbonyl complex **3c** on the solid phase. Immediately after the addition of the precursor complex $[(CH_3CN)_3Mo(CO)_3]$ to the swollen



Figure 2. UV/Vis/NIR spectra of soluble (top) and immobilised complexes (bottom).

resin 3a at room temperature the colour of the resin changes to deep blue, indicating formation of the immobilised complex while the solution remains yellow—the colour of the starting material—demonstrating the stability of the resinlinker-ligand bonds.

As mild reaction conditions are used, formation of $[(\eta^{6} \text{polystyrene})\text{Mo}(\text{CO})_{3}]^{[21]}$ is effectively prevented as evidenced by IR spectroscopy of resin **3b** and by a blank experiment using unmodified polystyrene and $[(\text{CH}_{3}\text{CN})_{3}\text{Mo}(\text{CO})_{3}]$ at ambient temperature.

Stirring resin **3b** in CH_2Cl_2 for 7 h gives the immobilised tetracarbonyl complex **3c** (according to colour and IR spectroscopy) in close analogy to the model reactions in solution (Scheme 5)—even the rate is similar (90% yield at 6.4 h in solution). Ligand substitution reactions were therefore performed in pure toluene, which is a suitable solvent for both the resin and the ligands L (Scheme 4).

Bubbling carbon monoxide through a suspension of **3b** in toluene causes a gradual colour change of the polymer from blue to purple, giving resin 3c while the solution remained colourless. The isonitrile ligand CNtBu and the phosphane ligand PPh₃ do not cause a significant colour change of the resin. The formation of the corresponding immobilised complexes 3d and 3e is evidenced by spectroscopic means (see below). In all cases the toluene solution remained colourless, indicating the stability of the resin-linker and linker-ligand bonds under these conditions. From the resinbound tetracarbonyl complex 3c the seven-coordinate complex 3f is formed by reaction with phenyltin trichloride (analogous to Scheme 6) again under colour change of the resin from purple to orange-red and CO evolution. In this case-unlike the solution model complex-the silyl linkage remains intact, probably because of the higher stability of the diisopropyl silvlether compared with that of the trimethyl silvl group.

The easy formation of the immobilised complexes indicates that the metal moieties on the resin are accessible even for bulky ligands such as PPh₃ and PhSnCl₃ so that all discussed reactions on the solid phase proceed similarly to their solution counterparts.

The stability of the different resin bound complexes 3b-3e towards ligand exchange follows the same trend of increasing lability as the soluble complexes $CNtBu < CO < PPh_3 < CH_3CN$ as shown by IR and ³¹P NMR spectroscopy and cleavage from the resin (see below).

Solid-phase chemistry, resin characterisation: Unlike the pure organic resin **3a** with solely covalent bonds between main group elements, mass spectrometry of the metal complex functionalised resins 3b-3f did not prove useful as the complexes decompose prior to resin depolymerisation.^[18] Also elemental analysis has proven unreliable for metal containing resins as has been shown previously.^[22]

For characterising the bulk material TG and DSC was used: DSC shows a shift of the peak maximum to lower temperature $(9-13 \,^{\circ}\text{C};$ see Table 4) compared with that of **3a** and the TG experiments show multistep weight losses starting around $100 \,^{\circ}\text{C}$, indicating the lower thermal stability of **3b-3e** compared with resin **3a**. The swelling of the bulk material is slightly lower than that of the organic polymer **3a** but this might only reflect the increased mass of the polymer. Again THF and toluene are the most suitable solvents for these resins.

The most useful spectroscopic method for studying the metal complex functionalised resins 3b-3f is IR spectroscopy (Table 4), especially in combination with a comparison to the spectra of the corresponding soluble complexes. In all cases the IR spectra (carbonyl region) of the immobilised complexes closely resemble those of the model complexes in solution (Figure 3). The similarity of corresponding spectra is



Figure 3. IR spectra of selected soluble and immobilised complex pairs (top: 2c/3c; middle: 2d/3d; bottom: 1f/3f). Bands marked with an asterisk belong to vibrations of the polystyrene.

much less when comparing the solid-state spectra indicting a "solution-like" environment for the immobilised complexes.

In addition to the strong carbonyl absorptions of the metal complexes the characteristic absorptions of the ligand-functionalised polymer are also seen around 2727, 1627, and 1584 cm⁻¹ (Table 4). For resin **3d** the $\tilde{\nu}_{CN}$ of the coordinated isonitrile ligand is observed at 2112 cm⁻¹.

In the diffuse reflectance spectra of the resins 3b-3f the MLCT bands are observed in the visible region (Table 4). Compared with their solution counterparts all absorption

bands are shifted to lower energy by $1500-3000 \,\mathrm{cm}^{-1}$ but preserve the trend $\lambda_{\max}(\mathbf{b}) > \lambda_{\max}(\mathbf{d}) \approx \lambda_{\max}(\mathbf{e}) > \lambda_{\max}(\mathbf{c}) > \lambda_{\max}(\mathbf{f})$ (Tables 3 and 4, Figure 2). The linear correlation between $\tilde{\nu}_{\max}$ of the soluble and the immobilised complexes and ligands is very good $\tilde{\nu}_{\max}(\text{immobilised molecules}) = \tilde{\nu}_{\max}(\text{soluble molecules}) \times 0.83+924 \,\mathrm{cm}^{-1}$ ($\mathbf{R}^2 = 0.9801$; see Supporting Information).

The gel-phase ¹³C NMR spectra of the polymers 3c-3fshow signals due to the linker and the attached ligand (Table 4). For 3d-3f signals due to the monodentate ligands CN*t*Bu, PPh₃, and PhSnCl₃ are also seen. In the gel-phase ³¹P NMR spectrum of 3e the signal of coordinated phosphane ligand appears at $\delta = 40$ as a broad signal (Figure 4). The half



Figure 4. ³¹P NMR spectra of **2e** (in CD₂Cl₂) and **3e** (in[D₈]THF).

width at half height is 250 times higher than in the ³¹P NMR spectra of the soluble complexes **1e** and **2e**, indicating that this signal indeed corresponds to a polymer-bound species with much lower mobility.

An attempt to observe the ¹¹⁹Sn signal of **3f** at room temperature failed. At -70 °C a broad signal occurs around $\delta = 50$. Its half width at half height is 35 times larger than that of the soluble analogue **1 f**, again indicating the observation of a polymer-bound species.

Attempts to obtain reliable ⁹⁵Mo NMR spectra of the resin bound complexes failed because of the low sensitivity of the ⁹⁵Mo nucleus and the significant "baseline ringing" in these experiments.^[15a]

In summary TG, DSC, and elemental analyses are not especially helpful in characterising the polymer-bound complexes as most of the measured quantities arise from the polymer backbone and not from the complexes of interest. In



Figure 5. ¹¹⁹Sn NMR spectra of 1 f and 3 f (in[D₈]THF).

contrast IR, diffuse reflectance, and NMR spectroscopy combine specificity for the metal complexes with the required sensitivity.

Solid-phase chemistry, cleavage: After having characterised the polymer-bound complexes directly on the solid support, we obtained indirect proof of identity through the possibility of mild cleavage of the complexes from the insoluble polymer (Scheme 9). Treating resins 3c-3f with two equivalents of



Scheme 9. Cleavage of the complexes from solid support.

- 2929

tetra-*n*-butylammonium fluoride in CH₂Cl₂ gives deeply coloured solutions of the deprotonated complexes. After washing the resins and concentrating the filtrates, careful acidification (at low pH the complexes start to decompose) of the solutions with acetic acid gives solutions of the respective complexes as evidenced by UV/Vis/NIR and IR spectroscopy in 30-40% yield over all steps based on the loading of **3a**. In all cases only the desired complexes are isolated with no indication of side products.

The IR spectra of the remaining slightly coloured resins prove the absence of carbonyl groups confirming quantitative cleavage from the resin. Additionally, mass spectrometry of the resins (Scheme 9) confirms the absence of attached ligands and the presence of silyl fluoride groups on the polymer.

Conclusion

Inorganic chemistry on the solid organic polystyrene phase is similar to solution chemistry of the investigated substitution and oxidative elimination reactions. Resin-bound ligands and complexes can be prepared and analysed and are readily cleaved from the solid support. Ligand substitution reactions and oxidative eliminations proceed somewhat more slowly on solid support compared to the homogeneous reaction. Care has to be taken when choosing the optimal solvent. The relative stability trend of the polymer-bound complexes is identical to that of the solution analogues. IR, DRS-UV, and NMR prove valuable to monitor the solid-phase reaction and to characterise the resin bound complexes. Finally, the investigated cleavage conditions are mild enough to obtain the free complexes from the solid support without major decomposition. With this knowledge it should now be possible to prepare inorganic and organometallic complexes on the solid phase, which might be accessible only with difficulty or even not at all in the homogeneous phase. Additionally, this methodology might be useful for combinatorial catalyst libraries when the catalysts must be tested in solution and therefore be cleaved from the support.

Experimental Section

Polystyrene (2% DVB), brominated polystyrene (2% DVB, 2.5 mmol Brg⁻¹), Cl₂Si(*i*Pr)₂, *n*BuLi in toluene (2.5 M), diisopropylethylamine, (dimethylamino)pyridine (DMAP), and *n*Bu₄NF · 3H₂O (TBAF) were purchased from Fluka, phenyltin trichloride from Strem. The polystyrene resins were washed prior to use with CCl₄, acetone, acetone/H₂O (2:1), acetone, benzene, MeOH, CH₂Cl₂, and diethyl ether and dried under vacuum. Unless noted otherwise, all manipulations were carried out under argon by means of standard Schlenk techniques. All solvents were dried by standard methods and distilled under argon prior to use. All other reagents were used as received from commercial sources. For the solid-phase reactions a flask with a nitrogen inlet and a coarse porosity fritted glass filter that allows addition and removal of reagents and solvents without exposure of the resin to the atmosphere was used.

NMR spectra were obtained on a Bruker Avance DPX 200 (200.15 MHz (¹H), 50.323 MHz (¹³C), 81.015 MHz (³¹P), 13.029 MHz (⁹⁵Mo), 74.630 MHz (¹¹⁹Sn)) at 303 K if not noted otherwise; chemical shifts (δ) are reported in ppm with respect to residual solvent peaks as internal standards: [D₈]THF (¹H: δ = 1.73; ¹³C: δ = 25.5), CD₃CN (¹H: δ = 1.93; ¹³C: δ = 1.3), CD₂Cl₂

(¹H: $\delta = 5.32$; ¹³C: $\delta = 53.8$); [d₆]DMSO (¹H: $\delta = 2.50$; ¹³C: $\delta = 39.4$). ³¹P chemical shifts (d) in ppm with respect to 85 % H_3PO_4 (31P: $\delta\!=\!0),$ 95Mo chemical shifts (δ) in ppm with respect to 1M Na₂MoO₄ in D₂O (⁹⁵Mo: δ = 0) and ¹¹⁹Sn chemical shifts (δ) in ppm with respect to SnMe₄ in THF (¹¹⁹Sn: $\delta = 0$) as external standards. IR spectra were recorded on a BioRad Excalibur FTS 3000 spectrometer using caesium iodide disks or CaF2 cells. UV/Vis/NIR spectra were recorded on a Perkin Elmer Lambda 19 in 0.2 cm cells (Hellma, suprasil). DRS spectra were measured with the same instrument using the Perkin Elmer integrating sphere and polytetrafluoroethylene (PTFE) as reference. Mass spectra were recorded on a Finnigan MAT 8400 spectrometer, electron energy 70 eV (EI), 4-nitrobenzyl alcohol matrix (FAB). Elemental analyses were performed by the microanalytical laboratory of the Organic Chemistry Department, University of Heidelberg. Cyclic voltammetry was performed on a Metrohm "Universal Meßund Titriergefäß", Metrohm GC electrode RDE 628, platinum electrode, SCE electrode, Princeton Applied Research potentiostat Model 273; substrate concentration 10-3 M in 0.1 M nBu₄NPF₆/CH₂Cl₂ or in 0.1 M nBu₄NPF₆/CH₃CN. All potentials are given relative to that of SCE. Differential scanning calorimetry measurements were carried out on a Mettler DSC 30, heating rate 10 K min⁻¹, under argon from 30-600 °C. Thermogravimetric measurements were carried out on a Mettler TC 15, heating rate 10 K min-1 under argon from 30-800 °C. Swelling measurements were performed by weighing a known amount of resin into a calibrated NMR tube and adding excess solvent. The volume of the swollen resin was measured after 3 h.

4-[(Pyridine-2-ylmethylene)amino]phenol (1a): Pyridine-2-carbaldehyde (15 g, 140.2 mmol), p-aminophenol (15.3 g, 140.2 mmol), MgSO₄ (8 g) and ethyl acetate (120 mL) were heated under reflux for 2 h. The solvent was removed under reduced pressure and the yellow residue was taken up in water and extracted with *n*-butanol $(3 \times)$. The alcohol was removed under reduced pressure and the yellow solid residue washed with water $(3 \times)$ and diethyl ether $(3 \times)$ and dried in vacuo, yielding **1a** (25 g, 126 mmol; 90%). ¹H NMR (200 MHz, [D₆]DMSO): $\delta = 6.86$ (d, ³J_{H,H} = 8.7 Hz, 2H; H^{3.5}), 7.32 $(d, {}^{3}J_{H,H} = 8.7 \text{ Hz}, 2 \text{ H}; \text{H}^{2.6}), 7.49 \text{ (pt, 1H; H}^{11}), 7.93 \text{ (pt, 1H; H}^{10}), 8.14 \text{ (d,})$ $J_{\rm H,H} = 7.8$ Hz, 1H; H⁹), 8.62 (s, 1H; H⁷), 8.70 (d, $J_{\rm H,H} = 4.6$ Hz, 1H; H¹²), 9.63 (s, 1 H; OH); ¹³C NMR (50 MHz, $[D_6]DMSO$): $\delta = 116.7$ (s; C^{3,5}), 121.6 (s; C⁹), 123.8 (s; C^{2,6}), 125.9 (s; C¹¹), 137.7 (s; C¹⁰), 142.4 (s; C¹), 150.4 (s; C12), 155.5 (s; C4), 157.8 (s; C7), 158.0 (s; C8) All signals were assigned on the basis of DEPT, CH-COSY and NOESY experiments. The conformations in solution are transoid as no cross peaks between H7 and H9 (numbering scheme shown in Scheme 2) are observed in the NOESY spectra; IR (CsI): $\tilde{\nu} = 3040 - 2472$ (m, br, OH, CH), 1626 (m), 1601 (m), 1581 (s), 1510 (s) cm⁻¹; UV/Vis (CH₂Cl₂): λ_{max} (ε) = 286 (15560), 340 nm (19740 m⁻¹ cm⁻¹); MS (EI, 70 eV): m/z (%): 198 (100) [M]⁺, 181 (2) $[M^+ - OH]^+$, 171 (45) $[M - HCN]^+$, 154 (3) $[M - OH - HCN]^+$, 79 (26) $[C_5H_5N]^+$; elemental analysis (%) calcd for $C_{12}H_{10}N_2O$ (198.22): C 72.71, H 5.08, N 14.13; found: C 72.62, H 5.09, N 14.08.

Pyridine-2-ylmethylene-(4-trimethylsilanyloxyphenyl)amine (2a): A suspension of 1a (2.97 g, 15 mmol) in THF (30 mL) was cooled to 0 °C. NEt₃ (1.57 g, 15.5 mmol) was added through a cannula. After the mixture had been stirred for 10 min, TMSCl (1.68 g, 15.5 mmol) was added and the suspension was stirred for 1 h at 0°C and 1 h at 25°C. The solvent was removed under reduced pressure and the yellow residue dissolved in diethyl ether. The yellow solution was filtered and the solvent removed under reduced pressure to give 2a as an orange-yellow oil (3.53 g, 87%). ¹H NMR (200 MHz, CD₂Cl₂): $\delta = 0.38$ (s, 9 H; CH₃), 6.99 (d, ³J_{H,H} = 8.8 Hz, 2H; $H^{3,5}$), 7.36 (d, ${}^{3}J_{HH} = 8.8$ Hz, 2H; $H^{2,6}$), 7.40 (pt, 1H; H^{11}), 7.84 (pt, 1H; H^{10}), 8.28 (d, $J_{H,H} = 8.0$ Hz, 1 H; H^9), 8.71 (s, 1 H; H^7), 8.74 (d, $J_{H,H} = 4.6$ Hz, 1 H; H¹²); ¹³C NMR (50 MHz, CD₂Cl₂): $\delta = 0.37$ (s; CH₃), 121.1 (s; C^{3,5}), 121.6 (s; C⁹), 123.0 (s; C^{2,6}), 125.2 (s; C¹¹), 136.9 (s; C¹⁰), 144.9 (s; C¹), 149.9 (s; C¹²), 155.1 (s; C⁴), 155.5 (s; C⁸), 159.1 (s; C⁷); IR (CsI): $\tilde{\nu} = 3060$ (w, CH_{Ar}), 2960 (m, CH₃), 1627 (m), 1597 (w), 1583 (s), 1503 (vs) cm⁻¹; UV/Vis $(CH_2Cl_2): \lambda_{max} (\epsilon) = 285 (21810), 337 \text{ nm} (23230 \text{ M}^{-1} \text{ cm}^{-1}); \text{ MS} (EI, 70 \text{ eV}):$ m/z (%): 270 (100) $[M]^+$, 269 (70) $[M-H]^+$, 255 (12) $[M-CH_3]^+$; elemental analysis (%) calcd for C15H18N2OSi (270.41): C 66.62, H 6.71, N 10.36; found: C 66.52, H 6.67, N 10.59.

fac-Tricarbonyl(2a)(acetonitrile)molybdenum(0) (2b): $[(CH_3CN)_3Mo(CO)_3]$ (303 mg, 1 mmol) was treated with 2a (270 mg, 1 mmol) in CH₃CN (20 mL) to give an intensely turquoise-coloured solution. After stirring for 3 h at room temperature the solution was concentrated to about 5 mL and petroleum ether (PE) 40/60 (40–50 mL) was added. After 24 h the

coloured solution was removed with a cannula and the dark residue was washed with PE 40/60 and dried in vacuo to give **2b** as a purple powder (417 mg; 85%). The reaction can also be performed on a 10 mmol scale. MS (FAB): m/z (%): 452 (78) $[M - CH_3CN]^+$, 424 (20) $[M - CH_3CN - CO]^+$, 396 (60) $[M - CH_3CN - 2 CO]^+$, 368 (100) $[M - CH_3CN - 3 CO]^+$; elemental analysis (%) calcd for $C_{20}H_{21}N_3O_4$ SiMo (491.43): C 48.68, H 4.29, N 8.52; found: C 48.30, H 4.33, N 8.50.

cis-Tetracarbonyl(2a)molybdenum(0) (2c): Carbon monoxide was bubbled into a solution of 2b (491 mg, 1 mmol) in CH₂Cl₂ (20 mL) until the IR spectrum showed only the presence of 2c resulting in a purple solution. The reaction mixture was stirred under a CO atmosphere for 1 h. Removal of the solvent in vacuo left 2c as a purple powder (477 mg; 100%). MS (FAB): m/z (%): 480 (45) $[M]^+$, 452 (100) $[M - CO]^+$, 424 (20) $[M - 2 CO]^+$, 396 (30) $[M - 3 CO]^+$, 368 (40) $[M - 4 CO]^+$; elemental analysis (%) calcd for C₁₉H₁₈N₂O₅SiMo (478.39): C 47.70, H 3.79, N 5.86; found: C 47.30, H 3.87, N 6.07.

fac-**Tricarbonyl(2a)**(*tert*-**butyl isonitrile)molybdenum(0) (2d)**: *tert*-Butyl isonitrile (0.11 mL, 1 mmol) was added to a solution of **2b** (491 mg, 1 mmol) in CH₂Cl₂ (20 mL) to give a deep blue solution. After stirring for 3 h at room temperature the solution was concentrated to about 5 mL and PE 40/60 (40–50 mL) was added. After 24 h the coloured solution was removed with a cannula and the dark blue residue was washed with PE 40/60 (and dried in vacuo to give **2d** as a blue powder (470 mg; 88%). MS (FAB): *m/z* (%): 535 (10) [*M*]+, 507 (50) [*M* – CO]+, 479 (18) [*M* – 2 CO]+, 451 (30) [*M* – 3 CO]+, 368 (100) [*M* – 3 CO – CN*t*Bu]+; elemental analysis (%) calcd for C₂₃H₂₇N₃O₄SiMo (533.51): C 51.78, H 5.10, N 7.88; found: C 50.55, H 4.86, N 7.78.

fac-**Tricarbonyl(2 a)(triphenylphosphane)molybdenum(0) (2 e)**: Triphenylphosphane (262 mg, 1 mmol) was added to a solution of **2b** (491 mg, 1 mmol) in CH₂Cl₂ (20 mL), giving a deep blue solution. After stirring for 3 h at room temperature the solution was concentrated to about 5 mL and PE 40/60 (40–50 mL) was added. After 24 h the coloured solution was removed with a cannula and the dark blue residue was washed with PE 40/60 and dried in vacuo to give **2e** as a blue powder (640 mg, 90%). MS (FAB): *m/z* (%): 714 (33) [*M*]⁺, 686 (80) [*M* – CO]⁺, 658 (30) [*M* – 2 CO]⁺, 630 (100) [*M* – 3 CO]⁺; elemental analysis (%) calcd for C₃₆H₃₃N₂O₄SiPMo (712.67): C 60.67, H 4.66, N 3.93, P 4.35; found: C 60.40, H 4.75, N 4.01, P 4.30.

fac-Tricarbonyl(1a)(acetonitrile)molybdenum(0) (1b): The preparation was similar to that of 2b but 1a (198 mg, 1 mmol) was used. The washing solution was almost colourless. Yield: 385 mg (92%). MS (FAB): m/z (%): 380 (100) $[M - CH_3CN]^+$, 352 (24) $[M - CH_3CN - CO]^+$, 324 (46) $[M - CH_3CN - 2 \text{ CO}]^+$; elemental analysis (%) calcd for $C_{17}H_{13}N_3O_4MO$ (419.25): calcd: C 48.70, H 3.13, N 10.02; found: C 48.08, H 3.15, N 9.87.

cis-**Tetracarbonyl(1a)molybdenum(0) (1 c)**: The preparation was similar to that of **2 c** but **1 b** (419 mg, 1 mmol) was used instead of **2 b**. Yield: 405 mg (100%). MS (FAB): m/z (%): 408 (47) $[M]^+$, 380 (100) $[M - CO]^+$; elemental analysis (%) calcd for $C_{16}H_{10}N_2O_5Mo$ (406.21): C 47.31, H 2.48, N 6.90; found C 46.90, H 2.80, N 6.85.

fac-Tricarbonyl(1a)(*tert*-butyl isonitrile)molybdenum(0) (1d): The preparation was similar to that of 2d but 1b (419 mg, 1 mmol) was used instead of 2b. Yield: 435 mg (94%). MS (FAB): m/z (%): 463 (65) $[M]^+$, 435 (100) $[M - CO]^+$; elemental analysis (%) calcd for $C_{20}H_{19}N_3O_4Mo$ (461.33): C 52.07, H 4.15, N 9.11; found: C 51.37, H 4.12, N 9.00.

fac-Tricarbonyl(1a)(triphenylphosphane)molybdenum(0) (1e): The preparation was similar to that of 2e but 1b (419 mg, 1 mmol) was used instead of 2b. Yield: 430 mg (93%). Recrystallisation from CH₂Cl₂/Et₂O afforded dark blue crystals. MS (FAB): m/z (%): 642 (48) $[M]^+$, 614 (80) $[M - CO]^+$, 586 (100) $[M - 2 \text{ CO}]^+$, 557 (91) $[M - 3 \text{ CO} - \text{H}]^+$; elemental analysis (%) calcd for C₃₃H₂₅N₂O₄PMo (640.49) · 0.5 Et₂O: C 62.05, H 4.46, N 4.13, P 4.57; found: C 61.73, H 4.53, N 4.23, P 4.52.

 μ -Chloro-(dichlorophenyltin)–(1a)tricarbonyl molybdenum(II) (1 f): Phenyltin trichloride (606 mg, 2 mmol) was added with syringe to a solution of 1 c (406 mg, 1 mmol) in acetone (20 mL). The solution was stirred for 5 h at room temperature until IR spectroscopy indicated completeness of the reaction. The colour turned from deep purple to orange-red. The solvent was removed under reduced pressure and the residue was washed with diethyl ether (6 × 20 mL) and dried to give a yellow-orange powder (550 mg; 81%). Recrystallisation from THF/Et₂O afforded orange-coloured crystals. MS (FAB): m/z (%): 645 (51) $[M - Cl]^+$, 561 (24) [M - Cl -

3 CO]⁺, 380 (100) $[M - \text{SnCl}_2\text{Ph}]^+$; elemental analysis (%) calcd for C₂₁H₁₅N₂O₄SnCl₃Mo (680.35) · 1.0 Et₂O: C 39.80, H 3.34, N 3.71; found: C 39.48, H 3.43, N 3.77.

Reaction of [(CH₃CN)₃Mo(CO)₃] with polystyrene: [(CH₃CN)₃Mo(CO)₃] (455 mg, 1.5 mmol) dissolved in CH₃CN was added to polystyrene (1 g; 2 % DVB) swollen in CH₃CN/toluene (1:2). The suspension was slowly stirred at room temperature for 3 h. The solvent was removed by filtration and the resin was washed repeatedly with CH₃CN, toluene, and Et₂O until the washings were colourless. The IR spectrum (absence of v_{CO} bands) indicated no formation of [(polystyrene)Mo(CO)₃].^[21]

Resin 3a: The resin was prepared from brominated polystyrene (2 % DVB, 2.5 mmol Br g⁻¹) according to a published procedure^[18] using the ligand **1a** to give a yellow-coloured polymer. MS (EI, 70 eV, 407 °C): *m/z* (%): 104 (100) $[C_8H_8]^+$, 329 (11) $[C_8H_7SiOL + H]^+$, 343 (43) $[C_8H_7SiMeOL]^+$, 372 (84) $[C_8H_7Si(iPr)OL]^+$, 415 (69) $[C_8H_7Si(iPr)_2OL]^+$; elemental analysis (%) calcd for $(C_8H_8)_{20}(C_{26}H_{30}SiON_2)_3(C_{15}H_{24}SiO)_7$: C 81.32, H 8.23, N 1.66; found C 78.93, H 7.38, N 1.56; which corresponds to 0.56 mmol **1a** per gram polymer. Uptake of CoCl₂ corresponds to 0.52 mmol per gram polymer.

Uptake of CoCl₂: A standard solution of CoCl₂ in THF was treated with a weighed amount of resin **3a**. From the decrease of the absorbances at 590 and 683 nm of the solution the loading was calculated by extrapolation to $t \rightarrow \infty$.

Resin 3b: $[(CH_3CN)_3Mo(CO)_3]$ (303 mg, 1.0 mmol) dissolved in CH₃CN (5 mL) was added to resin **3a** (1 g; corresponding to 0.56 mmol **1a**) swollen in toluene (5 mL). The suspension was allowed to stand at room temperature for 24 h with occasional stirring. The solvents were removed by filtration and the polymer was washed with CH₃CN (4 × 10 mL), toluene/CH₃CN (10 mL), toluene (2 × 10 mL), and finally with diethyl ether (2 × 10 mL). The blue resin was dried in vacuo to give a blue powder or was directly allowed to react with the ligands L (see below). Elemental analysis (%) calcd for $(C_8H_8)_{20}(C_{31}H_{33}SiO_4N_3MO)_3(C_{15}H_{24}SiO)_7$: C 75.05, H 7.51, N 2.20; found: C 77.41, H 7.32, N 1.68.

Resin 3c: CO was bubbled for 15 min into a suspension of resin **3b** (starting from 1 g of **3a**, 0.56 mmol) swollen in toluene (6 mL). The blue resin turned purple. The suspension was allowed to stand at room temperature under CO for 24 h with occasional stirring. The solvents were removed by filtration and the polymer was washed with toluene (1×10 mL) and diethyl ether (2×10 mL). The resin was dried in vacuo giving a purple powder. Elemental analysis (%) calcd for (C_8H_8)₂₀($C_{30}H_{33}$ SiO₅N₂Mo)₃($C_{15}H_{24}$ SiO)₇: C 74.94, H 7.40, N 1.48; found: C 77.53, H 7.06, N 1.42.

Resin 3d: To resin **3b** (starting from 1 g of **3a**, 0.56 mmol) swollen in toluene (6 mL) CN*t*Bu (0.11 mL, 1.0 mmol) was added with a syringe. The suspension was allowed to stand at room temperature for 8 h with occasional stirring. The solvents were removed by filtration and the polymer was washed with toluene (5×10 mL) and diethyl ether (2×10 mL). The dark blue-green resin was dried in vacuo giving a dark blue-green powder. Elemental analysis (%) calcd for (C_8H_8)₂₀($C_{34}H_{39}$ SiO₄N₃-MO)₃($C_{15}H_{24}$ SiO)₇: C 75.28, H 7.66, N 2.15; found: C 78.61, H 7.45, N 1.66.

Resin 3e: PPh₃ (262 mg, 1.0 mmol) dissolved in toluene (3 mL) was added with a syringe to resin **3b** (starting from 1 g of **3a**, 0.56 mmol) swollen in toluene (3 mL). The suspension was allowed to stand at room temperature for 24 h with occasional stirring. The solvents were removed by filtration and the polymer was washed with toluene (5×10 mL) and diethyl ether (2×10 mL). The dark blue resin was dried in vacuo to give a dark blue powder. Elemental analysis (%) calcd for (C_8H_8)₂₀($C_{47}H_{45}SiO_4N_2P-MO$)₃($C_{15}H_{24}SiO_7$: C 76.28, H 7.30, N 1.31, P 1.46; found: C 76.71, H 6.85, N 1.29, P 0.87.

Resin 3 f: Phenyltin trichloride (303 mg, 1 mmol) was added with a syringe to resin **3 c** (starting from 1 g of **3 a**, 0.56 mmol) swollen in toluene (6 mL). The suspension was allowed to stand at room temperature for 24 h with occasional stirring. The solvents were removed by filtration and the polymer was washed with toluene $(4 \times 10 \text{ mL})$ and diethyl ether $(2 \times 10 \text{ mL})$. The resin was dried in vacuo giving an orange-red powder. Elemental analysis (%) calcd for $(C_8H_8)_{20}(C_{35}H_{36}SiO_4N_2MOSnCl_3)_3(C_{15}H_{24}-SiO)_7$: C 68.25, H 6.70, N 1.29; found: C 69.13, H 6.59, N 1.33.

Cleavage of the resin-bound complexes: Tetra-*n*-butylammonium fluoride (TBAF \cdot 3H₂O, 315 mg, 1 mmol) in CH₂Cl₂ (5 mL) was added to the appropriate resin (starting from 1 g of **3a**, 0.56 mmol) swollen in CH₂Cl₂ (3 mL). The suspension was stirred slowly for 12 h. The coloured solution was removed by filtration and the resin was washed thoroughly with

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CH₂Cl₂/Et₂O mixtures until the solutions remained colourless. The combined filtrates were dried in vacuo. Dissolving the dark residue in CH₂Cl₂ (100 mL) and careful acidification with degassed acetic acid (0.1 mL) gave intensely coloured solutions of the respective complexes. The yields were determined by UV/Vis spectrophotometry and the identities were confirmed by IR spectroscopy. Overall yields (metallation, substitution, cleavage, and acidification) based on the loading of **3a** were between 30 and 40 %. After cleavage, samples of the remaining resins were subjected to mass spectrometry, which showed complete removal of the complexes. MS (EI, 70 eV, 400 °C): m/z (%): 104 (100) [C₈H₈]⁺, 151 (18) [C₈H₇Si(*i*Pr)₂F]⁺.

Acknowledgements

This work was supported by the Deutsche Forschungsgemeinschaft and the Fonds der Chemischen Industrie. The permanent, generous support from Prof. Dr. G. Huttner is gratefully acknowledged.

- [1] F. Zaragoza Dörwald, *Organic Synthesis on Solid Phase*, Wiley-VCH, Weinheim, **2000**.
- [2] a) K. S. Schmidt, D. M. Filippov, N. J. Meeuwenoord, G. A. van der Marel, J. H. van Boom, B. Lippert, J. Reedijk, *Angew. Chem.* 2000, *112*, 383-385; *Angew. Chem. Int. Ed.* 2000, *39*, 375-377; b) S. I. Khan, A. E. Beilstein, M. W. Grinstaff, *Inorg. Chem.* 1999, *38*, 418-419; c) S. I. Khan, A. E. Beilstein, M. Sykora, G. D. Smith, X. Hu, M. W. Grinstaff, *Inorg. Chem.* 1999, *38*, 3922-3925; d) A. E. Beilstein, M. W. Grinstaff, *Chem. Commun.* 2000, 50-510.
- [3] M. S. Robillard, A. R. P. M. Valentijn, N. J. Meeuwenoord, G. A. van der Marel, J. H. van Boom, J. Reedijk, *Angew. Chem.* 2000, *112*, 3226–3229; *Angew. Chem. Int. Ed.* 2000, *39*, 3096–3099.
- [4] S. Tadesse, A. Bhandari, M. A. Gallop, J. Comb. Chem. 1999, 1, 184– 187.
- [5] a) N. E. Schore, S. D. Najdi, J. Am. Chem. Soc. 1990, 112, 441-442;
 b) A. C. Comely, S. E. Gibson, N. J. Hales, Chem. Commun. 2000, 305-306; c) A. C. Comely, S. E. Gibson, N. J. Hales, Chem. Commun. 1999, 2075-2076; d) S. E. Gibson, N. J. Hales, M. A. Peplow, Tetrahedron Lett. 1999, 40, 1417-1418; e) M. F. Semmelhack, G. Hilt, J. H. Colley, Tetrahedron Lett. 1998, 39, 7683-7686; f) N. Mensi, S. S. Isied, J. Am. Chem. Soc. 1987, 109, 7882-7884; g) B. A. Arbo, S. S. Isied, Int. J. Peptide Protein Res. 1993, 42, 138-154.
- [6] a) J. B. Arterburn, K. V. Rao, M. C. Perry, Angew. Chem. 2000, 112, 787-788; Angew. Chem. Int. Ed. 2000, 39, 771-772; b) F. Minutolo, J. A. Katzenellenbogen, Angew. Chem. 1999, 111, 1730-1732; Angew. Chem. Int. Ed. 1999, 38, 1617-1620.
- [7] a) B. Altava, M. I. Burguete, J. M. Fraile, J. I. García, S. V. Luis, J. A. Mayoral, M. J. Vicent, *Angew. Chem.* 2000, *112*, 1563–1566; *Angew. Chem. Int. Ed.* 2000, *39*, 1503–1506; b) R. Drake, R. Dunn, D. C. Sherrington, S. J. Thomson, *Chem. Commun.* 2000, 1931–1932; c) S. Tamagaki, R. J. Card, D. C. Neckers, *J. Am. Chem. Soc.* 1978, *100*, 6635–6639.
- [8] a) H. Miessner, K. Richter, J. Mol. Catal. A 1999, 146, 107-115; b) H.
 Miessner, J. Am. Chem. Soc. 1994, 116, 11522-11530; c) B. B. Kaul,

R. K. Hayes, T. A. George, J. Am. Chem. Soc. 1990, 112, 2002–2003;
d) G. Gubitosa, H. H. Brintzinger, J. Organomet. Chem. 1977, 140, 187–193;
e) G. Gubitosa, M. Boldt, H. H. Brintzinger, J. Am. Chem. Soc. 1977, 99, 5174–5175;
f) R. H. Grubbs, C. P. Lau, R. Cukier, C. H. Brubaker, J. Am. Chem. Soc. 1977, 99, 4517–4518;
g) W. D. Bonds, C. H. Brubaker, E. S. Chandrasekaran, C. Gibbons, R. H. Grubbs, L. C. Kroll, J. Am. Chem. Soc. 1975, 97, 2128–2132;
h) R. H. Grubbs, C. Gibbons, L. C. Kroll, W. D. Bonds, C. H. Brubaker, J. Am. Chem. Soc. 1975, 97, 2128–2132;
h) R. H. Grubbs, C. Gibbons, L. C. Kroll, W. D. Bonds, C. H. Brubaker, J. Am. Chem. Soc. 1973, 95, 2373–2375;
i) S. Dahmen, S. Bräse, Angew. Chem. 2000, 112, 3827–3830; Angew. Chem. Int. Ed. 2000, 39, 3681–3683.

- [9] S. Bräse, D. Enders, J. Köbberling, F. Avemaria, Angew. Chem. 1998, 110, 3614–3616; Angew. Chem. Int. Ed. 1998, 37, 3413–3415.
- [10] a) B. M. Trost, I. Hachiya, J. Am. Chem. Soc. 1998, 120, 1104–1105;
 b) B. M. Trost, S. Hildbrand, K. Dogra, J. Am. Chem. Soc. 1999, 121, 10416–10417;
 c) B. M. Trost, M. Lautens, J. Am. Chem. Soc. 1983, 105, 3343–3344;
 d) B. M. Trost, M. Lautens, J. Am. Chem. Soc. 1982, 104, 5543–5545;
 e) F. Glorius, A. Pfaltz, Org. Lett. 1999, 1, 141–144;
 f) N.-F. K. Kaiser, U. Bremberg, M. Larhed, C. Moberg, A. Hallberg, Angew. Chem. 2000, 112, 3742–3744; Angew. Chem. Int. Ed. 2000, 39, 3596–3598;
 g) M. P. T. Sjögren, H. Frisell, B. Åkermark, P.-O. Norrby, L. Eriksson, A. Vitagliano, Organometallics, 1997, 16, 942–950.
- [11] W. Kaim, S. Kohlmann, Inorg. Chem. 1987, 26, 68-77.
- [12] a) H. Behrens, E. Lindner, G. Lehnert, J. Organomet. Chem. 1970, 22, 439-448; b) A. Srivastavsa, A. K. Shrimal, S. C. Srivastavsa, Polyhedron 1994, 13, 975-982; c) B. Bildstein, M. Malaun, H. Kopacka, M. Fontani, P. Zanello, Inorg. Chim. Acta 2000, 300-302, 16-22; d) L. W. Houk, G. R. Dobson, Inorg. Chem. 1966, 5, 2119-2123; e) M. N. Ackermann, W. G. Fairbrother, N. S. Amin, C. J. Deodene, C. M. Lamborg, P. T. Martin, J. Organomet. Chem. 1996, 523, 145-151; f) E. C. Alyea, V. K. Jain, Polyhedron 1995, 15, 1723-1730.
- [13] R. W. Balk, D. J. Stufkens, A. Oskam, *Inorg. Chim. Acta* 1978, 28, 133–143.
- [14] R. A. Grieves, J. Mason, Polyhedron 1986, 5, 415-421.
- [15] a) J. Malito, Ann. Rep. NMR Spectrosc. 1997, 33, 151–206; b) M. Minelli, W. J. Maley, Inorg. Chem. 1989, 28, 2954–2958.
- [16] S. Song, E. C. Alyea, Can. J. Chem. 1996, 74, 2304-2320.
- [17] a) A. Bell, R. A. Walton, J. Organomet. Chem. 1985, 290, 341–352;
 b) R. Kummer, W. A. G. Graham, Inorg. Chem. 1968, 7, 310–315;
 c) M. Elder, W. A. G. Graham, D. Hall, R. Kummer, J. Am. Chem. Soc. 1968, 90, 2189–2190;
 d) M. Panizo, M. Cano, J. Organomet. Chem. 1985, 287, 221–229.
- [18] K. Heinze, U. Winterhalter, T. Jannack, Chem. Eur. J. 2000, 6, 4203 4210.
- [19] a) J. T. Randolph, K. F. McClure, S. J. Danishefsky, J. Am. Chem. Soc.
 1995, 117, 5712-5719; b) S. J. Danishefsky, K. F. McClure, J. T. Randolph, R. B. Ruggeri, Science 1993, 206, 1307-1309; c) L. A. Thompson, F. L. Moore, Y.-C. Moon, J. A. Ellman, J. Org. Chem. 1998, 63, 2066-2067.
- [20] R. Santini, M. C. Griffith, M. Qi, *Tetrahedron Lett.* 1998, 39, 8951– 8954.
- [21] C. P. Tsonis, M. F. Farona, J. Organomet. Chem. 1976, 114, 293-298.
- [22] R. J. Card, D. C. Neckers, *Inorg. Chem.* 1978, 17, 2345–2349.

Received: December 20, 2000 [F2955]